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Sex in Plants Requires Thrust

NEWS [\(/cell-science/news\)](/cell-science/news)  Sep 25, 2018 | By Katherine Gombay for McGill University
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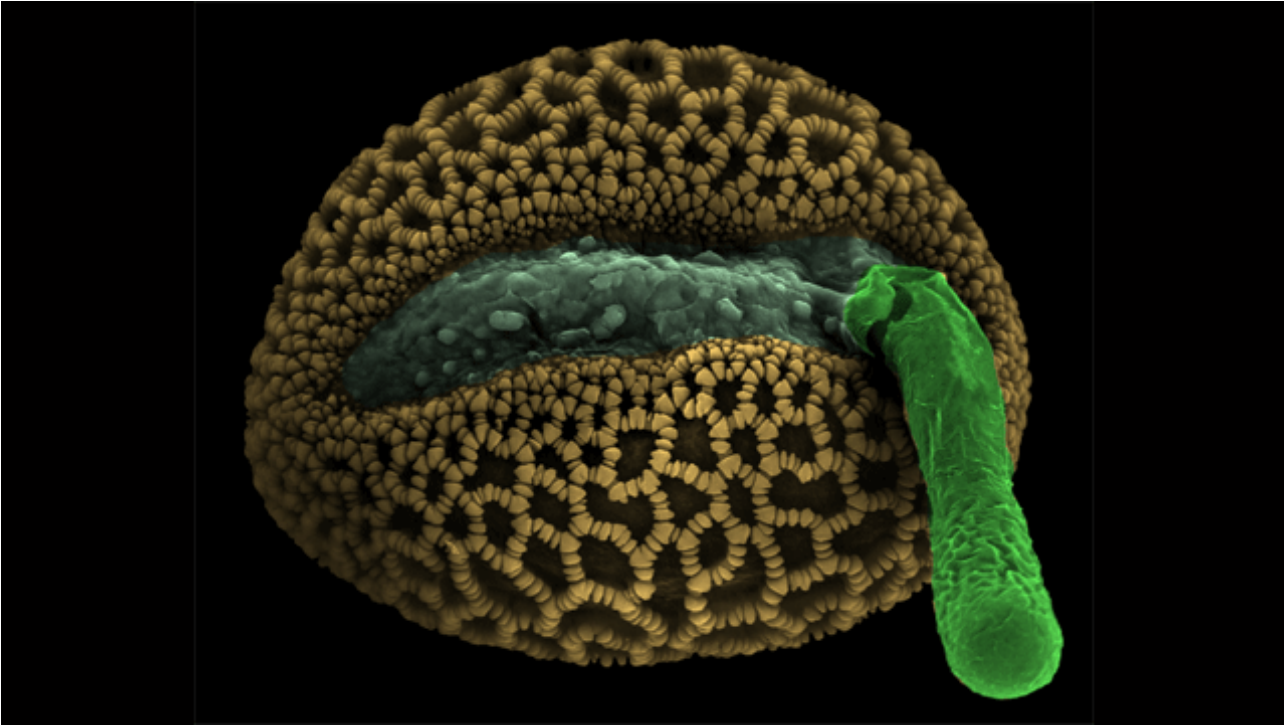


Photo credit: McGill University

Plant sex relies on a combination of prodding and a lot of communication and guidance, suggests a study published in the September 2018 issue of *Technology*.

It's a process that is fraught with challenges. The sperm, two of which are housed in each grain of pollen, are unable to move on their own and the egg cell is deeply embedded in the pistil (the female tissues of the flower).

So, to reach the egg the sperm rely on a pollen tube that extends into the pistil. These invasive tubes are the fastest growing cells in the plant kingdom, growing up to 1-2 cm (or 500x their original dimension) an hour, and can sometimes extend up to 30 cm, depending on the anatomy of the flower.

To fertilize the egg, the pollen tube (which is between 1/20 and 1/5 of the width of a human hair) has to navigate through a maze of tissue, no matter what obstacles it encounters.

The phenomenon is well-documented and known to require communication at cellular level with the female flower tissues, but relatively little is understood about the cell mechanics involved. So scientists from McGill and Concordia collaborated to look more closely at the growth force of individual pollen tubes using a microfluidic lab-on-a-chip.

"From a mechanical point of view, the process of pollen tube elongation is similar to that of a balloon catheter

used in angioplasty – forces are generated based on fluid under pressure,” explains Muthukumaran Packirisamy from Concordia University’s Department of Mechanical and Industrial Engineering. “So, we designed a microscopic cantilever with a gauge built-in that the pollen tubes had to forcefully push against in order to continue to elongate.”

Anja Geitmann, formerly at l’Université de Montréal who is now Canada Research Chair in McGill’s Department of Plant Science is the senior author on the paper. She adds:

“Thanks to the lab-on-a chip technology we were able to actually see and measure exactly what was going on within the pollen tube as it grew. We discovered that the water pressure and force that these tiny cells exert as they push through the plant tissue to reach their destination is equivalent to the air pressure we put in our car tires to keep them rolling...

What is even more exciting is that we found that when the pollen tube encounters an obstacle, it changes its growth pattern, suggesting that the cells are in some ways able to ‘feel’ and respond to the physical resistance in their environment. It’s very exciting to be able to see this process, and it leaves us with a lot of interesting questions ahead about male-female communication.”

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Reference:

Ghanbari, M., Packirisamy, M., & Geitmann, A. (2018). Measuring the growth force of invasive plant cells using Flexure integrated Lab-on-a-Chip (FiLoC). *Technology*, 1-9. doi:10.1142/s2339547818500061 (<https://www.worldscientific.com/doi/abs/10.1142/S2339547818500061?journalCode=technology>)

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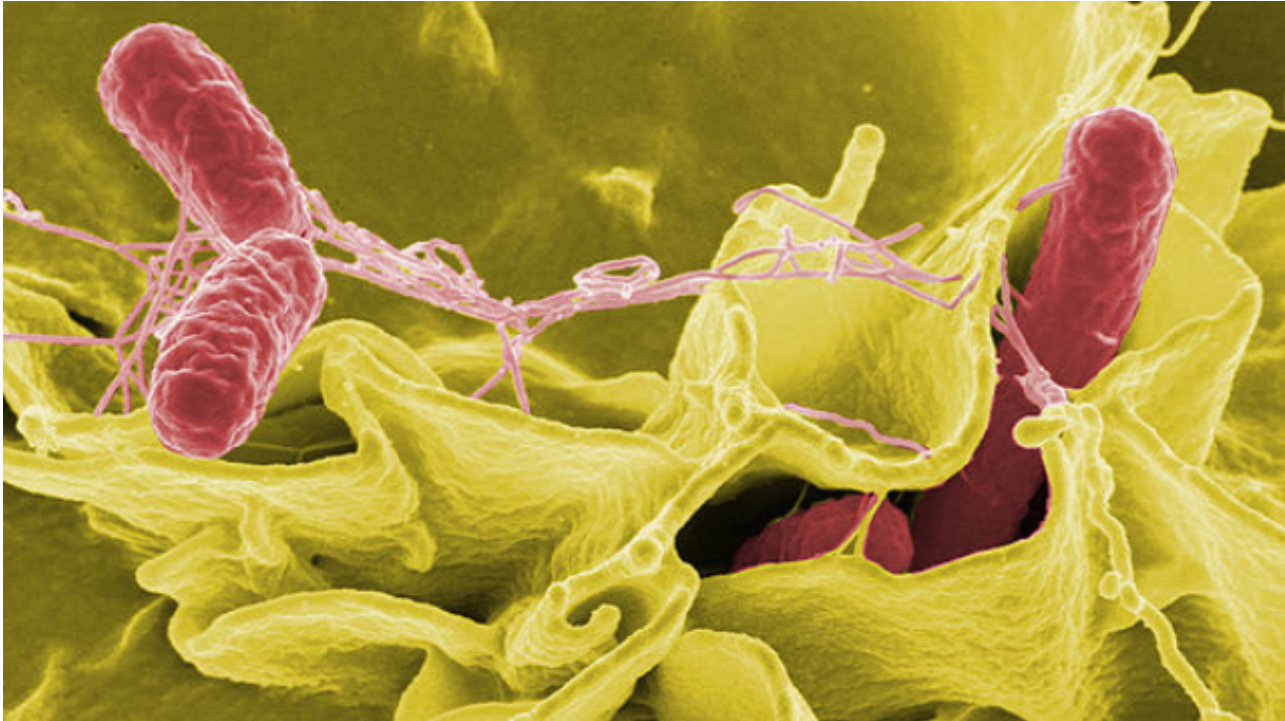
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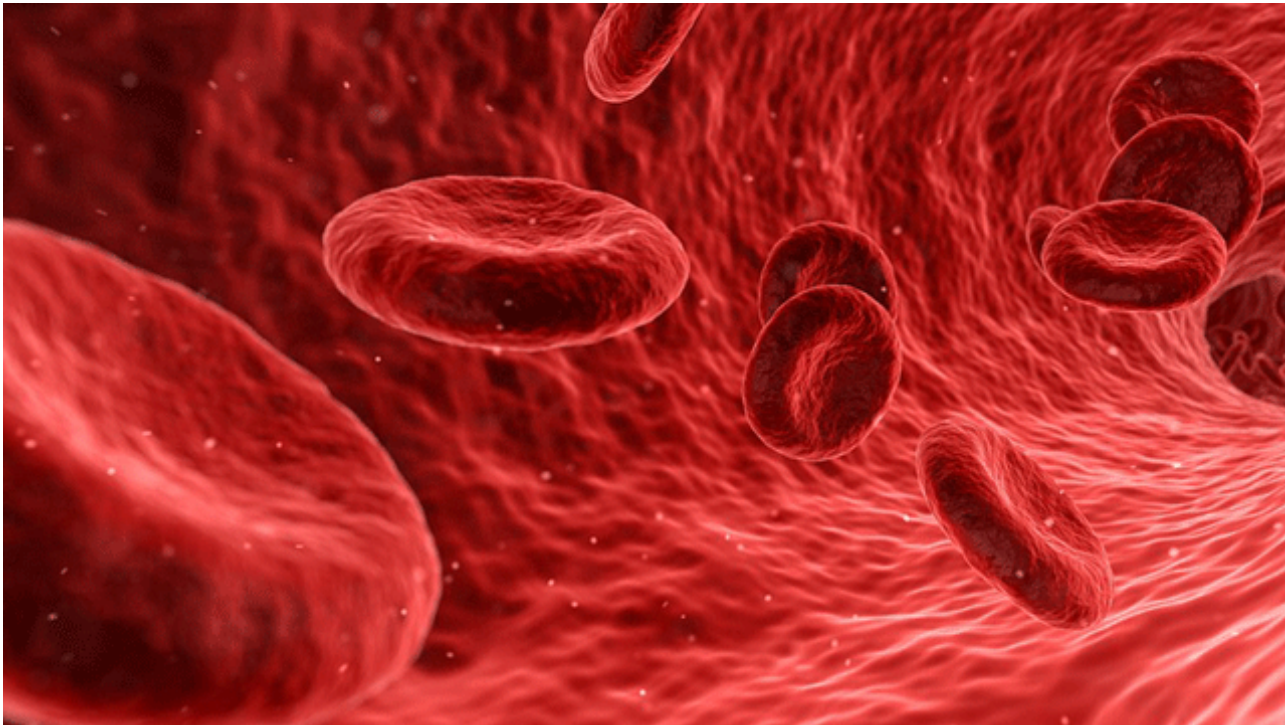
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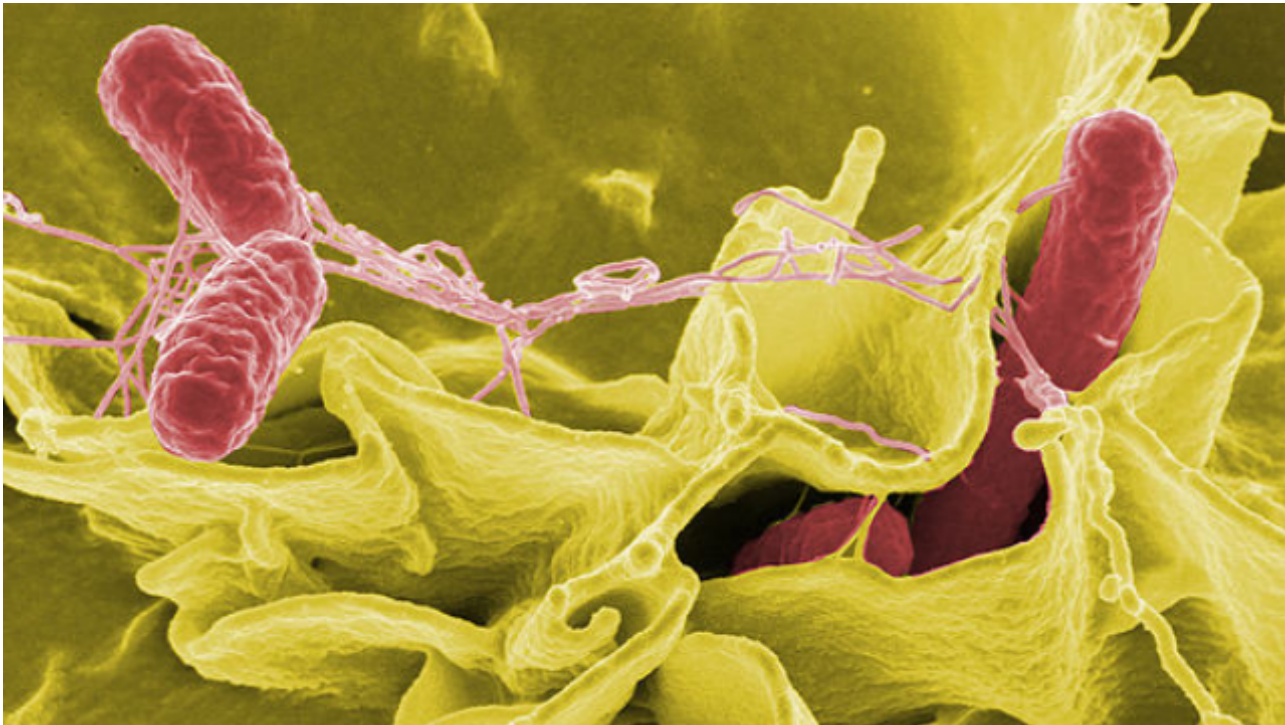
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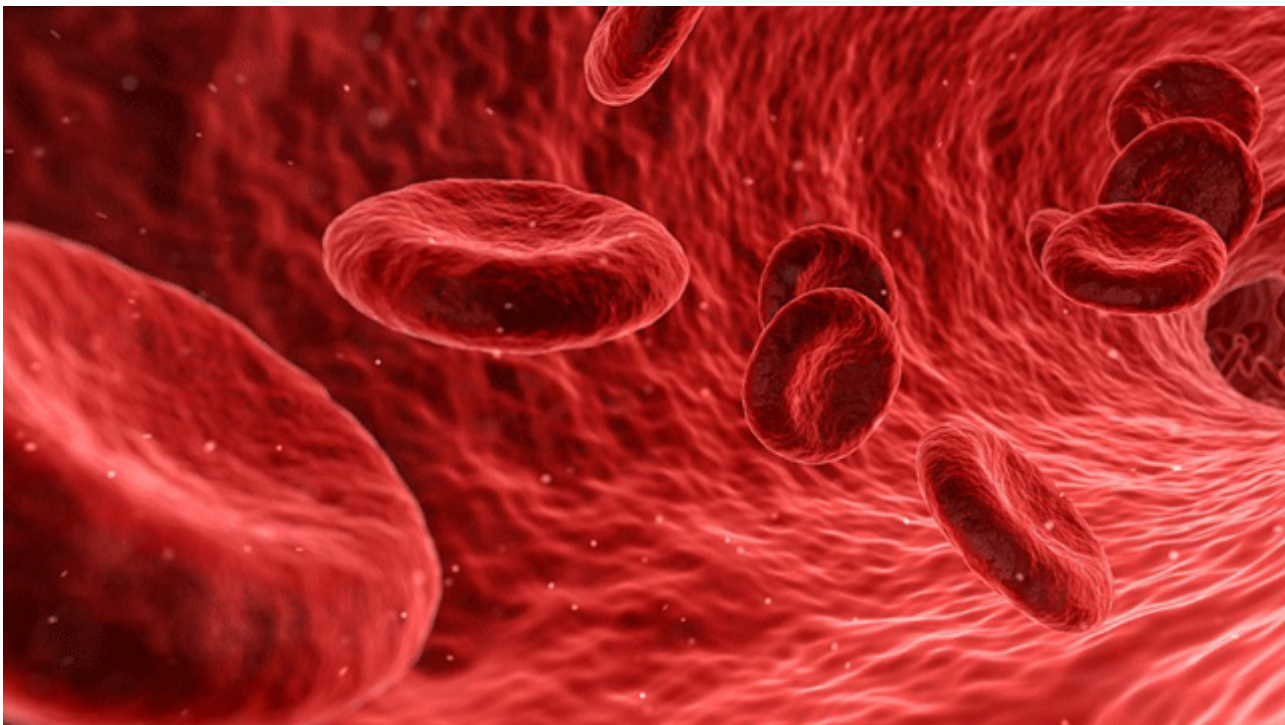
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M. Sayeed, Y. Zang, C. Moncada, T. Giardiello, D. Chimento, K. Abarca Heidemann
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Limerick, PA 19464

Introduction

Efficient analysis of Host Cell Protein (HCP) impurities is a crucial prerequisite for testing products in the drug manufacturing process. Generic CHO HCP assays are frequently employed early in biologic development when the process is still poorly defined, biologic manufacturers often migrate to a process-specific HCP assay in later stages of development. The generic HCP assay has utility supporting early process development workflow.

The quality of HCP antibody reagents can vary depending on antibody manufacturing conditions and robust characterization is therefore needed to ensure quality. HCP antibody reagents are most commonly evaluated via ELISA and orthogonal methods like one and two dimensional western blot assays (1D and 2D) where the sensitivity and coverage of the antibody are established. Appropriate coverage across a 2D-blot is demonstrated using 2 criteria. First the percent coverage is determined by identifying the number of detected protein species compared to the number of existing species shown on a corresponding 2D SDS-PAGE. The second criteria is quadrant analysis, that is used to demonstrate the proper detection of proteins having diverse sizes and isoelectric points. Well-developed HCP reagents show broad coverage that includes the low molecular weight (LMW) proteins. Here we evaluate and demonstrate improved coverage of a new generic CHO-HCP reagent developed at Rockland. Comparison of coverage to a leading commercial reagent was performed. We observed differences in both percent total coverage between each reagent, with a significant variance in the lower quadrants.

Antibody development


HCP type	CHO Proteins		
Ab Host	Rb		
Validation	ELISA	1D	2D
Coverage	%		

2D-Western Blot Coverage


CHO HCP proteins detected by a commercial and Rockland's anti-CHO HCP antibody on a membrane (WB) via HRP detection. Proteins are equated to the proteins separated on a separate SDS-gel and visualized by an high sensitivity in-gel protein stain.

A) Commercial generic Antibody


In-Gel Total Protein Stain (Orion)



Commercial Anti-CHO HCP Antibody Detection



Commercial Anti-CHO HCP Antibody Spot Designation in Quadrants



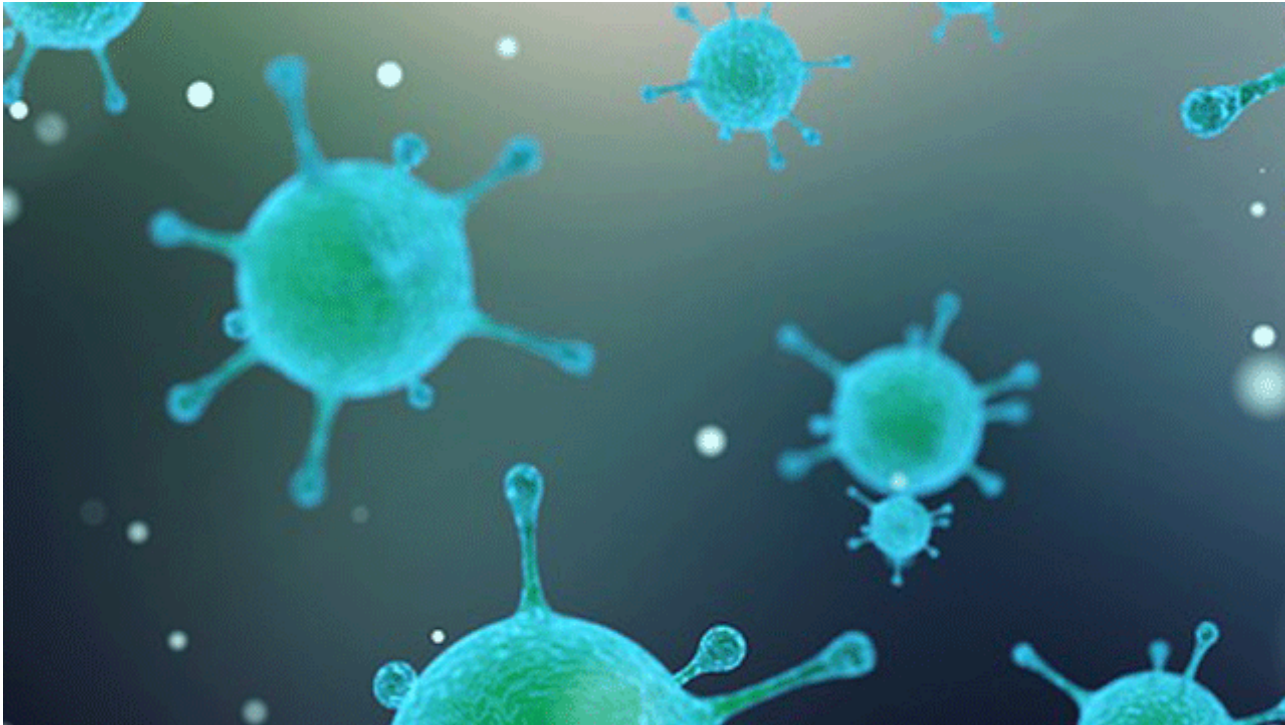
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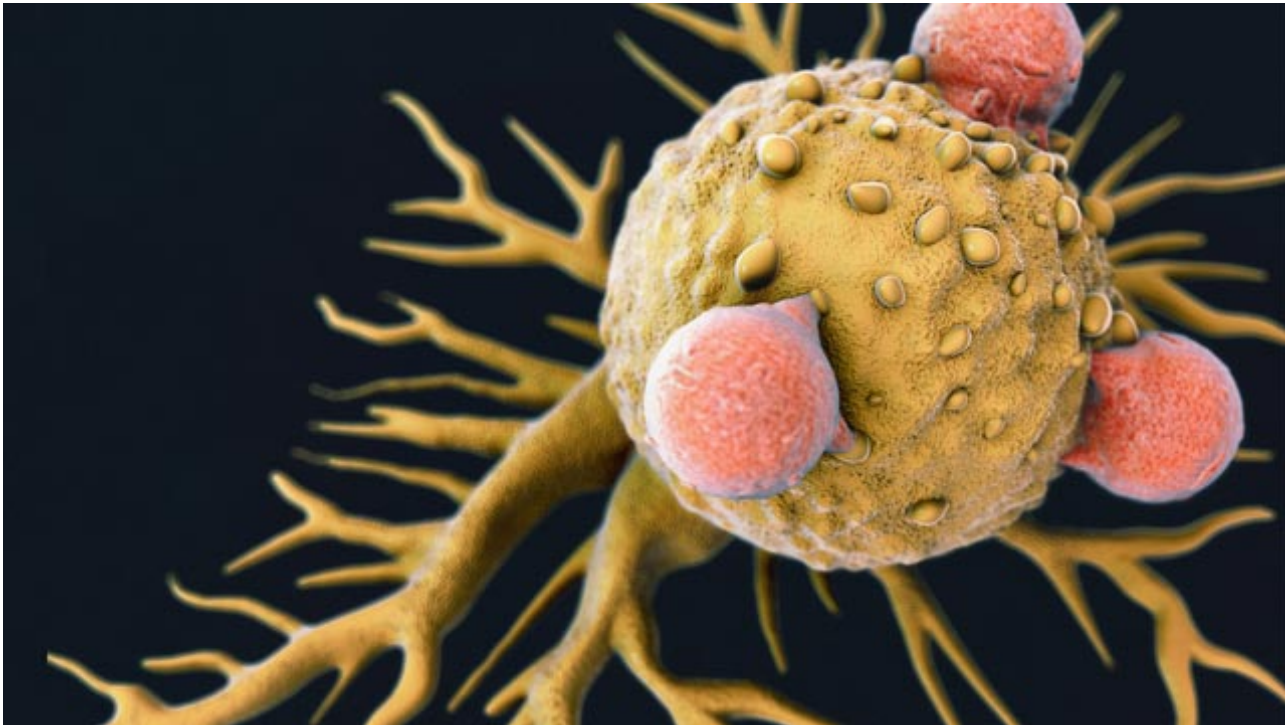
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